

**cDNA & Biotinylated cRNA Synthesis Reactions for GeneChips**

Date: \_\_\_\_\_

Samples:

Sample ID	Sample Descr.	RNA Input Vol

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C1	1 ug of mRNA Control	2 ul

Reagents/Materials needed:

Invitrogen Superscript Choice kit – includes:  
 5X 1<sup>st</sup> Strand Buffer      5X 2<sup>nd</sup> strand buffer  
 0.1M DTT                      DNA ligase (lot #: \_\_\_\_\_)  
 10mM dNTP                    DNA polymerase I (lot#: \_\_\_\_\_)  
 DEPC-H2O (variable)      RNase H (lot# \_\_\_\_\_)  
 SSII RT (variable: see table)

Table 1

Input RNA (ug)	SSRT (ul)
5-8	1
8.1-16	2
16.1-24	3
24.1-32	4
32.1-40	5

T7-(dT)24 primer (100 pmol/ul): 1 ul per sample  
 Optional: RNasin RNase Inhibitor (Promega)

**1. cDNA synthesis**

PCR PROGRAM = “ROB” => “CDNA”

ALL DONE ON ICE!! Keep enzymes in benchtop cooler at all times.

**STEP 1: Primer hybridization**

- Calculate amount of RT enzyme needed (see table above), **then** calculate H2O.
- Total Volume of STEP 1+2+3 = **20 ml**

								CI
Total RNA								2
DEPC-H2O								9
T7 Primer								1

- 70 C, 10 mins.  
 - Trx to ice, pulse spin

**STEP 2: Temperature adjustment**

								CI
5x 1 <sup>st</sup> St. Buff.	4	4	4	4	4	4	4	4
0.1M DTT	2	2	2	2	2	2	2	2
10mM dNTPs	1	1	1	1	1	1	1	1

- Add reagents  
 - 42C, 2mins

**STEP 3: First strand synthesis**

RT used: supplier : \_\_\_\_\_, lot number: \_\_\_\_\_ Rnase Inh. Lot (Promega Rnasin): \_\_\_\_\_

								CI
SSII RT								1
RNase Inh.								

- 42 C, 1 hr.  
 - Start:  
 - End:  
 -Trx to ice. Pulse spin, 4C

**STEP 4: Second strand synthesis**

							<b>CI</b>
<i>DEPC-H2O</i>	91	91	91	91	91	91	91
<i>5x 2<sup>nd</sup> S. Buff</i>	30	30	30	30	30	30	30
<i>10mM dNTPs</i>	3	3	3	3	3	3	3
<i>DNA Ligase</i>	1	1	1	1	1	1	1
<i>DNA Pol I</i>	4	4	4	4	4	4	4
<i>Rnase H</i>	1	1	1	1	1	1	1

- Mix gently. Pulse spin down, 4 C.
- 16 C, 2 hr.  
Start:  
End:
- Pulse spin down, 4C

**STEP 5: T4 DNA Polymerase**

							<b>CI</b>
<i>T4 DNA Pol</i>	2	2	2	2	2	2	2

- 16 C, 5 min

- Add 10 ul EDTA (0.5M, pH 8) to each vial to quench.
- Mix, place on ice, trx to new 1.5 ml eppendorf tubes. (If stopping here, store at -20.)

**Clean-up of (ds) cDNA:**

- Spin down PLG-I heavy tubes, 30 sec, max rpm, RT.
- To each sample, add equal vol. Phenol-chloroform-isoamyl alcohol (25:24:1) = **162 ul**. Vortex quickly.
- Trx to PLG-I tubes. (**DO NOT VORTEX.**) Spin 2 mins, max rpm, RT.
- Upper phase to new tube, labeled "G", dated: \_\_\_\_\_  
 Volumes:    1:     3:     5:  
                   2:     4:     C1:

**Immediate Ethanol Precipitation:**

							<b>CI</b>
<i>CDNA</i>							
<i>0.5 V 7.5 M Rnase-free NH<sub>4</sub>OAc</i>							
<i>2.5 V 100% EtOH</i>							
<i>Glycogen</i>	2	2	2	2	2	2	2
<i>Pellet Paint</i>	1	1	1	1	1	1	1

- Spin, 20 minutes, max rpm, 4 C.
- Wash 2x 500 ul 70% EtOH. 5 mins each spin, max rpm, 4 C
- Dry pellet. (Pipette, syringe, swab, air 5').
- Resuspend pellet in 12 ul DEPC-H2O

